NUCLEIC ACIDS HYBRIDIZATION

Nucleic Acids Hybridization Modern Applications

Edited by

ANTON BUZDIN

Shemyakin-Ovchinnikov, Institute of Bioorganic Chemistry, Moscow, Russia

and

SERGEY LUKYANOV

Shemyakin-Ovchinnikov, Institute of Bioorganic Chemistry, Moscow, Russia



A C.I.P. Catalogue record for this book is available from the Library of Congress.
ISBN 978-1-4020-6039-7 (HB) ISBN 978-1-4020-6040-3 (e-book)
Published by Springer, P.O. Box 17, 3300 AA Dordrecht, The Netherlands.
www.springer.com
Printed on acid-free paper
Cover idea is the courtesy of Dr. Lilia M. Ganova-Raeva
All Rights Reserved © 2007 Springer No part of this work may be reproduced, stored in a retrieval system, or transmitted in any form or by any means, electronic, mechanical, photocopying, microfilming, recording or otherwise, without written permission from the Publisher, with the exception of any material supplied specifically for the purpose of being entered and executed on a computer system, for exclusive use by the purchaser of the work.

CONTENTS

Contributors	xi
Acknowledgments	XV
Preface	vii
Nucleic Acids Hybridization: Potentials and Limitations	1
1. Introduction	2
2. Cloning the Differences: Subtractive Hybridization	4
2.1 Birth of a Method	4
2.2 PCR-assisted Subtractive Hybridization	8
2.3 First Worldwide Success: Representational	
Differences Analysis	10
2.4 Further Improvements: Suppression Subtractive	
Hybridization, Polymerase Chain Reaction Suppression	
Effect, and Normalization of cDNA Libraries	13
2.5 Covalently Hybridized Subtraction, Primer Extension	
Enrichment Reaction, and Other Promising Approaches	
in Subtractive Hybridization	18
3. Finding Common DNA: Coincidence Cloning	20
4. Hybridization in Solution for the Recovery of	
	22
5. Conclusions	24
References	25

vi Contents

2.	Selective Suppression of Polymerase Chain Reaction	
		29
	Sergey A. Lukyanov, Konstantin A. Lukyanov, Nadezhda G. Gurskaya,	
	Ekaterina A. Bogdanova, Anton A. Buzdin	
	1. Introduction	30
		30
		32
		33
		33
	5.1 Construction of cDNA Libraries from a Small Amount	-
		35
		36
		40
	ϵ	40
		41
	ϵ	42
	1	42
		43
		45
	References	48
3.	Suppression Subtractive Hybridization	53
•	Sergey A. Lukyanov, Denis Rebrikov, Anton A. Buzdin	
		54
		55
		57
	1	59
		59 59
		61
	ϵ	
		61
	2	62
		63
	ϵ	63
		63
		65
	6.1 Preparation of Subtracted cDNA or Genomic	
		65
	6.2 Mirror Orientation Selection	70
		74
		74
		77
		83

Contents vii

4.	Stem-Loop Oligonucleotides as Hybridization Probes and Their Practical Use in Molecular Biology and Biomedicine <i>Anton A. Buzdin, Sergey A. Lukyanov</i>	85
	1. Introduction	86 87 92 93 94
5.	Normalization of cDNA Libraries	97
	1. Introduction	98
	1.1 Normalized cDNA Libraries: What are they needed for?	98
	1.2 Evaluation of cDNA Library Normalization Efficacy1.3 Basic Approaches to Generate Normalized	100
	cDNA Libraries	100
	2. Methods of cDNA Normalization	102
	2.1 cDNA Normalization by Means of Hydroxyapatite	
	Column Chromatography	
	Libraries Using DNA Immobilization on a Solid Support	104
	2.3 Normalization of Full-Length cDNA with the	105
	use of Biotinylated RNA	103
	2.4 Normalization of Fragmented cDNA by Means of Selective Amplification	107
	2.5 cDNA Normalization Using Frequent-Cutter	107
	Restriction Enzymes	100
	2.6 Normalization of Full-Length-Enriched cDNA	10)
	with Duplex-Specific Nuclease (DSN Normalization)	109
	3. cDNA Preparation for DSN Normalization	
	3.1 RNA Requirements	
	3.2 cDNA Synthesis	
	3.3 cDNA Purification	113
	4. DSN Normalization Protocol	
	4.1 Materials	113
	4.2 cDNA Precipitation	114
	4.3 Hybridization	
	4.4 DSN Treatment	
	4.5 First Amplification of Normalized cDNA	
	4.6 Preliminary Analysis of the Normalization Results	
	4.7 Second Amplification of Normalized cDNA	

viii Contents

6.	Primer Extension Enrichment Reaction (PEER)	
	and Other Methods for Difference Screening	125
	Lilia M. Ganova-Raeva	
	1. Introduction	127
	2. Primer Extension Enrichment Reaction	128
	2.1 Method Outline	128
	2.2 Discussion	132
	2.3 PEER Protocol	135
	3. Other Subtraction and Hybridization Based Methods	
	for Difference Screening	
	3.1 Differential Screening	
	3.2 Subtractive Hybridization	139
		140
	3.4 Differential Display	
	3.5 AFLP, SAGE/CAGE, GSTs, and DARFA	
	3.6 Representational Differences Analysis	
	3.7 SPAD–RDA	146
	3.8 Enzymatic Degradation Subtractions (EDS, LCS,	
	DSC, NSC, UDG/USA, and CODE)	
	3.9 Suppression Subtraction Hybridization	150
	3.10 Selective Amplification Via Biotin and	
	Restriction-Mediated Enrichment	153
	3.11 DNA Enrichment by Allele-Specific	
	Hybridization	154
	3.12 Methods Combining the use of SSH	
	and Microarrays	
	3.13 Conclusions	
	References	157
7.	Subtractive Hybridization with Covalently	
	ϵ	167
	Shi-Lung Lin, Donald Chang, Joseph D. Miller, Shao-Yao Ying	
	1. Introduction	
	2. Subtractive Hybridization Methods	
	3. Covalent Modification	171
	4. Subtractive Hybridization with Covalently Modified	
	Subtracters	
	5. Applications	
	6. Protocols	
	6.1 Preparation of Subtracter and Tester DNA Libraries	
	6.2 Covalent Modification of Subtracter DNAs	
	6.3 Subtractive Hybridization and CHS–PCR Amplification	181

Contents ix

	6.4 Covalent Binding Efficiency and Subtractive	100
	Stringency of CHS	
	6.5 Identification of Genomic Deletion Using CHS	
	References	185
8.	Coincidence Cloning: Robust Technique for Isolation	
	of Common Sequences	187
	Anton A. Buzdin	
	1. Introduction	
	2. Cloning Selection of Heteroduplexes	191
	3. Physical Separation of Hybrids	
	4. PCR-only-based Approaches	193
	5. Future Prospects	202
	6. Protocols	
	6.1 Cloning Similarities in Genomic DNAs	202
	6.2 Cloning and Presice Mapping of Transcribed	20.4
	Repetitive Elements	204
	6.3 Finding Methylated or Unmethylated CpGs in Large Genomic Contigs	207
	References	207
	References	209
9.	DNA Hybridization in Solution for Mutation Detection	211
	Anton A. Buzdin	
	1. Introduction	
	2. Chemical Approaches	
	3. Enzymatic Approaches	
	3.1 Nuclease-Based Mutation Scanning	
	3.2 Allele-Specific PCR-Based Approaches	229
	3.3 Other Enzymatic Approaches for Mutation	220
	Scanning	
	4. Physical Approaches	
	5. Bioinformatical Approaches	
	References	230
10.	Current Attempts to Improve the Specificity of Nucleic Acids	
	Hybridization	241
	Anton A. Buzdin	
	1. Introduction	
	2. Improving Hybridization Kinetics	
	2.1 Simplification of Hybridizing Mixtures	
	2.2 Chemical Modifications	
	3. Improving Selection of Perfectly Matched Hybrids	249

x Contents

4. Protocols4.1 Targeted Genomic Difference Analysis4.2 Using Competitor DNA to Decrease	255
the Background of Genomic Repeats	
4.3 Mispaired DNA Rejection	
References	262
11. Concepts on Microarray Design for Genome	
and Transcriptome Analyses	265
Helder I. Nakaya, Eduardo M. Reis, Sergio Verjovski-Almeida	
1. Building a Microarray Chip	266
1.1 Spotted DNA Microarrays	268
1.2 <i>In situ</i> Synthesis	271
2. Selecting the Probes	
2.1 Gene-Oriented Arrays	278
2.2 Epigenomic Microarrays	
2.3 Tiling Arrays	
3. Specific Question, Specific Chip	280
3.1 Transcriptional Profiling	
3.2 Comparative Genome Hybridization	
3.3 Alternative Splicing	
3.4 Transcriptome Annotation	
3.5 Small MicroRNA Profiling	
3.6 Methylation Pattern	
3.7 ChIP-Chip	
3.8 Genotyping	
3.9 Intronic Transcription	
4. Conclusions	
References	
Index	309

CONTRIBUTORS

Ekaterina A. Bogdanova

Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry Moscow 117997, Miklukho-Maklaya 16/10, Russia

Anton A. Buzdin

Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry Moscow 117997, Miklukho-Maklaya 16/10, Russia

Nadezhda G. Gurskava

Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry Moscow 117997, Miklukho-Maklaya 16/10, Russia

Konstantin A. Lukyanov

Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry Moscow 117997, Miklukho-Maklaya 16/10, Russia

Sergey A. Lukyanov

Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry Moscow 117997, Miklukho-Maklaya 16/10, Russia

Denis Rebrikov

Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry Moscow 117997, Miklukho-Maklaya 16/10, Russia xii Contributors

Dmitry A. Shagin

Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry Moscow 117997, Miklukho-Maklaya 16/10, Russia

Alex S. Shcheglov

Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry Moscow 117997, Miklukho-Maklaya 16/10, Russia

Pavel A. Zhulidov

Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry Moscow 117997, Miklukho-Maklaya 16/10, Russia

Donald Chang

Department of Cell and Neurobiology, Keck School of Medicine, University of Southern California, 1333 San Pablo Street, BMT-403, Los Angeles, CA 90033, USA

Shi-Lung Lin

Department of Cell and Neurobiology, Keck School of Medicine, University of Southern California, 1333 San Pablo Street, BMT-403, Los Angeles, CA 90033, USA

Joseph Miller

Department of Cell and Neurobiology, Keck School of Medicine, University of Southern California, 1333 San Pablo Street, BMT-403, Los Angeles, CA 90033, USA

Shao-Yao Ying

Department of Cell and Neurobiology, Keck School of Medicine, University of Southern California, 1333 San Pablo Street, BMT-403, Los Angeles, CA 90033, USA

Lilia M. Ganova-Raeva

Centers for Disease Control and Prevention Division of Viral Hepatitis 1600 Clifton Rd. NE, MS A-33 Atlanta, Georgia 30329, USA Contributors xiii

Helder I. Nakaya

Departamento de Bioquimica, Instituto de Quimica, Universidade de São Paulo, 05508-900 São Paulo, SP, Brazil

Eduardo M. Reis

Departamento de Bioquimica, Instituto de Quimica, Universidade de São Paulo, 05508-900 São Paulo, SP, Brazil

Sergio Verjovski-Almeida

Departamento de Bioquimica, Instituto de Quimica, Universidade de São Paulo, 05508-900 São Paulo, SP, Brazil

ACKNOWLEDGMENTS

Anton A. Buzdin:

Many thanks to Professor Eugene D. Sverdlov for his fruitful discussion, innovative ideas, and overall support of this project. Thanks to my colleagues, friends, and family members for their help, patience, and understanding. A. Buzdin was funded by the Molecular and Cellular Biology Program of the Presidium of the Russian Academy of Sciences, by the personal grant from the President of the Russian Federation, and by Russian Foundation for Basic Research grants Nos. 05-04-48682-a, 2006.20034.

Sergey A. Lukyanov, Alex Shcheglov, Pavel Zhulidov, Ekaterina Bogdanova, Dmitry Shagin:

Work on this book was supported by the Molecular and Cellular Biology Program of the Russian Academy of Sciences and Evorogen JSC (Moscow, Russia).

Lilia M. Ganova-Raeva:

Special thanks to Dr. Y. Khudyakov who contributed most to the PEER backbone idea and has been relentlessly resourceful, helpful, and patient throughout the development of the method. Thanks to Dr. H. Fields in whose lab the PEER testing was initiated. Thanks to Dr. X. Zhang for great help with the library screenings and to Dr. F. Cao for introducing better enzymes in the protocol.

Shi-Lung Lin, Donald Chang, Joseph D. Miller, Shao-Yao Ying:

This study was supported by NIH/NCI grant CA-85722. Rhw CHS technology is the property of University of Southern California and protected by US patent numbers, 5,928,872 and 6,130,040.

Helder I. Nakaya, Eduardo M. Reis, Sergio Verjovski-Almeida:

The work in the authors' laboratory was supported by grants and fellowships from Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP), and Conselho Nacional do Desenvolvimento Científico e Tecnológico (CNPq), Brazil.

PREFACE

Watson-Crick hybridization of complementary sequences in nucleic acids is one of the most important fundamental processes necessary for molecular recognition in vivo, as well as for nucleic acid identification and isolation in vitro. This book is devoted to a large family of in vitro DNA hybridization-based experimental techniques. A wide spectrum of experimental tasks covered by these approaches includes finding differential sequences in both genomic DNAs and mRNAs, genome walking, multiplex PCR, cDNA library construction starting from minute amount of total RNA, rapid amplification of cDNA 5'- and 3'ends, effective smoothing of the concentrations of rare and abundant transcripts in cDNA libraries, recovery of promoter active repeats and differentially methylated genomic DNA, identification of common sequences in genomic or cDNA sources, new gene mapping, finding evolutionary conserved DNA and both single-nucleotide and extended mutation discovery, or large-scale monitoring. Several approaches, such as microarray hybridization, have become extremely popular tools for specialists in biochemistry and biomedicine, whereas the potential of many other advantageous techniques seems to be underestimated now.

Analysis of differential gene expression requires application of global approaches that represent a leading tool in postgenomic studies and include transcriptome and proteome analysis, as well as methods allowing population-wide sequence and functional polymorphism analysis. Central to these new technologies are DNA chips designed for quantitative and qualitative uses. Although they are very useful and widely distributed, many popular DNA microarray techniques share a number of shortcomings:

1. The analysis is limited by a number of cDNAs/synthetic oligonucleotides applied on the chip. This number is usually significantly lower than the total

gene quantity of the organisms under study. It creates, therefore, a problem, that many genes escape such an analysis.

- 2. General transcriptome-wide chip techniques in their actual state hardly distinguish between different gene splice forms.
- 3. The expression of genes transcribed at low levels cannot be detected by using standard microarray approaches.
- 4. cDNA-based chips do not differentiate between many gene family members and/or between many transcripts containing repetitive DNA.
- 5. Microarray chips lack many natural RNAi, cDNAs, or synthetic oligonucleotides and, therefore, cannot be used for the comprehensive study of gene expression regulation at the level of RNA interference by small interfering RNAs.

However, most of these concerns can be effectively addressed by using specific variants of microchip technology, thus making microarrays a truly universal technique (see Chapter 11). Probably, the most important disadvantage of closed systems such as microarrays is that they require preliminary genomic sequence information in order to identify differentially expressed transcripts.

Open systems have the flexibility of identifying uncatalogued sequences. Related experimental techniques, based on DNA hybridization in solution, may be advantageous for many applications, starting from representative cDNA library construction for expressed sequence tag (EST) sequencing, to the identification of evolutionary conserved sequences, differentially expressed genes, or genomic deletions. Unique characteristics of many such techniques make them powerful competitors for well-known approaches that are appreciated world-wide like microarray and competitive genomic hybridizations. Nucleic acid hybridization in solution has few general advantages over hybridization with solid carrier-immobilized nucleic acids: faster hybridization kinetics, better discrimination of proper hybrids, and their availability for further PCR amplification and cloning. Among such in-solution hybridization methods, subtractive hybridization is undoubtedly the most popular technique.

Many techniques have a low efficiency of identifying rare genes that are differentially expressed. This problem is exacerbated when the change in expression level of rare transcripts is small. Since genes expressed at low levels also play a role in establishing differentiated phenotypes, their identification is essential for a complete mechanistic understanding of cellular changes. The major advantage of subtractive hybridization lies in the ability to identify differentially transcribed genes, irrespective of the level of expression, in the absence of sequence information. In addition to preparation of differential cDNA libraries, subtractive hybridization is also extremely useful for identification of genomic DNA differences.

Coincidence cloning, on the contrary, makes it possible to identify sequences which are common for all samples under comparison; cDNA normalization, which is used for smoothing of rare and frequent transcript content in cDNA libraries, may be extremely useful for representative EST library construction. Moreover, several techniques deal with the large-scale DNA polymorphism recovery, including identification of single nucleotide polymorphisms.

Preface xix

The international team of the authors of this book has tried both to elucidate the current state of the art in hybridization techniques and to help the readers in choosing an appropriate method for performing an experiment in the most efficient way. Enclosed experimental protocols along with both comprehensive and detailed method descriptions make this truly universal book useful to all those interested in the modern life science methodologies.